Gel Electrophoresis (Agarose) Experiment Template

Structured Documentation for DNA Fragment Analysis — From Gel Casting Through Electrophoresis, UV Imaging, and Per-Lane Band Scoring

Applies to

- Academic and molecular-biology research laboratories
- Biotechnology and life-sciences R&D groups
- Cloning, genotyping, and nucleic-acid QC teams
- Contract research organizations (CROs)

Relevant compliance & integrity frameworks

- **FDA 21 CFR Part 11** — Electronic records and signatures
- **GLP** — Good Laboratory Practice for preclinical documentation
- **Data integrity (ALCOA+)** — Attributable, contemporaneous, defensible records
- **NIH / NSF** — Reproducibility and data-management requirements

LabLynx, Inc.
3350 Riverwood Pkwy SE, Suite 1900, Atlanta, GA 30339
866.522.5969 | elabeln.com | info@lablynx.com

 © 2026 LabLynx, Inc. — Public information

1. Executive Summary

Gel electrophoresis is the moment a molecular-biology result becomes visible, and its output is decision data: the bands in each lane, sized against a ladder, are what tell you whether a digest worked, which clone carries the right insert, and which sample advances to sequencing or isolation. But a gel is only as trustworthy as the run and the interpretation behind it. A band called “the right size” by eye, with no ladder reference or recorded expected size, is not a weak result — it is an unverifiable one. And when gel images, lane layouts, expected sizes, and per-clone calls live scattered across a gel-doc drive, a lab-book sketch, and someone’s memory, results cannot be compared reliably across runs and clones — which is exactly what a cloning or screening project needs.

The ELabELN Gel Electrophoresis (Agarose) Experiment Template solves this by providing a pre-built, fully structured gel record that loads from the template library in a single click. It ships with a 10-step agarose protocol, validated buffer and ladder references, pre-wired voltage and run-time fields, and typed inputs for agarose concentration, buffer system, voltage, run time, stain, ladder identity, sample identity, expected fragment sizes, and per-lane observed bands. The agarose, buffer, ladder, and stain link to the ELabELN inventory, and the tamper-evident audit trail captures every gel run, every imaged result, and every clone-selection decision.

This white paper describes what the template contains, the gel-analysis use case it is designed to support, the upstream and downstream workflows it connects to, the benefits it delivers to the scientist, and how a laboratory can deploy it and begin running gels within days.

> **Key benefit**
> Labs that adopt the Gel Electrophoresis template stop rewriting the gel setup every run and lose no lane, expected size, or clone call between the imager and the notebook. Observed bands are scored against expected sizes per lane, every gel carries its run parameters and reagent lots, and outcomes become queryable across runs and clones — the structured, comparable foundation a cloning or screening project and defensible result provenance depend on.

 © 2026 LabLynx, Inc. — Public information

2. What Is Agarose Gel Electrophoresis, and Why Does Documentation Matter?

2.1 The Technique

Agarose gel electrophoresis separates nucleic-acid fragments by size. A gel is cast at a percentage chosen for the fragment range — lower for large fragments, higher for small ones — in a running buffer such as TAE or TBE. Samples mixed with a loading dye are pipetted into wells, a DNA ladder of known sizes is loaded alongside, and an electric field drives the negatively charged DNA through the gel toward the positive electrode; smaller fragments migrate faster and travel farther. A fluorescent stain such as SYBR Safe makes the DNA visible under UV or blue light, and the gel is imaged. Each band's size is read by comparing its migration to the ladder, and — for a diagnostic run — the observed bands are compared to the sizes expected from the construct, clone, or digest.

A band is only meaningful if the run and the reading support it: an appropriate agarose percentage and voltage, a ladder to size against, a clean image, and a recorded expectation to compare to. Weakness anywhere — a percentage that cannot resolve two close fragments, a run pushed too far, or a size guessed without a ladder — produces a result that looks conclusive and is not.

Because the bands decide which clone advances, which digest is confirmed, and which sample is isolated, and are compared across runs, **the gel image, the lane layout, the expected sizes, and the per-lane call are what make a gel trustworthy and comparable to the next run — which is exactly why they must live in one structured, queryable record rather than a folder of gel-doc images.**

2.2 The Documentation Problem

Recording gels across gel-doc drives, lab-book sketches, and memory creates a set of interconnected risks:

- The lane layout is sketched in a notebook, so which lane held which sample or clone is easily lost once the image is filed.
- The expected fragment sizes are not recorded, so an observed band is judged "about right" by eye instead of against a documented expectation.
- The gel image sits in a gel-doc export, disconnected from the samples, digest, and run parameters it depicts.
- Agarose percentage, buffer, voltage, and run time are rebuilt from memory, so an inconsistent run quietly makes band positions non-comparable.
- Ladder identity and stain are not captured, so a sizing discrepancy cannot be traced to a different ladder or a degraded stain.
- Per-lane pass/fail calls are remembered rather than recorded, so which clones passed cannot be queried later.
- Reagent lots are not captured against the run, so a run-to-run change cannot be traced to a reagent batch.

These are the gaps that make gel results hard to trust and compare, and they are the gaps this template was designed to close.

 © 2026 LabLynx, Inc. — Public information

3. Template Overview: What Ships with the Template

The ELabELN Gel Electrophoresis (Agarose) Experiment Template is a pre-populated experiment record available to all ELabELN subscribers directly from the template library. It ships configured for diagnostic restriction-digest verification of a pUC19-GFP cloning intermediate using an EcoRI + BamHI double digest, with a 10-step protocol covering gel casting through UV imaging and per-clone scoring. The default protocol assumes a 1.2% agarose gel in TAE buffer with SYBR Safe stain, 100 V for 50 minutes on a Mini-Sub Cell GT or equivalent rig, a 1 kb Plus DNA Ladder reference, and an 8-lane layout: ladder, six clones, and an undigested control. Every section is editable — the configuration is a tested baseline, not a locked format.

3.1 Template Sections at a Glance

The template populates the existing ELabELN experiment sections your lab already works in. Your team edits a working baseline instead of rewriting the gel setup from scratch every run.

Section	What it pre-populates
Main Text	The gel objective, sample-layout notes, and expected band pattern, authored in the TinyMCE rich-text editor, with expected fragment sizes inline and a link to the upstream digest.
Extra Fields	Ten structured, typed fields grouped as Gel, Run, and Scoring — agarose concentration, buffer system, voltage, run time, stain, ladder identity, sample identity, expected fragment sizes, per-lane observed bands, and per-lane score. Typed values make gel parameters queryable across runs.
Steps	A 10-step workflow checklist covering gel casting through imaging. Each tick is timestamped to the audit trail, and the imager run is auto-logged.
Compounds	The agarose, TAE buffer, SYBR Safe stain, DNA ladder, and loading dye pre-linked from the compound database, with lot numbers carried in automatically and the stain expiry shown inline.
Links & Resources	Pre-wired to the lab's Gel Electrophoresis SOP v2.4, the 1 kb Plus ladder reference image, and the digest setup for this run.
Storage & Files	4 °C stain working-stock storage, plus labelled attachment slots for gel images (TIF) and per-lane scoring sheets (CSV).

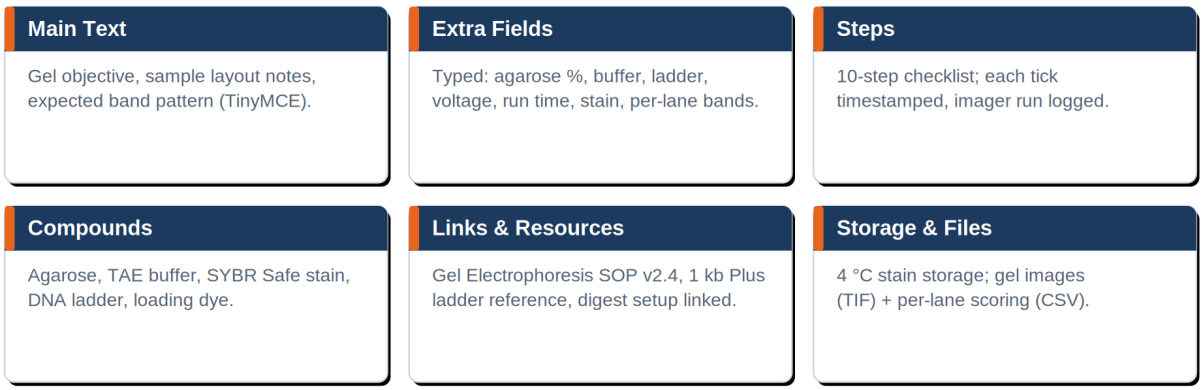
Main Text
Gel objective, sample layout notes, expected band pattern (TinyMCE).
Extra Fields
Typed: agarose %, buffer, ladder, voltage, run time, stain, per-lane bands.
Steps
10-step checklist; each tick timestamped, imager run logged.
Compounds
Agarose, TAE buffer, SYBR Safe stain, DNA ladder, loading dye.
Links & Resources
Gel Electrophoresis SOP v2.4, 1 kb Plus ladder reference, digest setup linked.
Storage & Files
4 °C stain storage; gel images (TIF) + per-lane scoring (CSV).

Figure 1. The six ELabELN experiment sections pre-populated by the Gel Electrophoresis (Agarose) template.

© 2026 LabLynx, Inc. — Public information

3.2 The 10-Step Protocol

The workflow checklist is the operational spine of the template. Each step maps to a discrete bench or analysis action and writes a timestamped, user-attributed entry to the audit trail when checked off. The imager run is auto-logged, and the UV imaging and band sizing are done with the gel-doc system, with the resulting bands and calls recorded as typed fields.

Step	Action	What it captures
1	Cast gel (1.2% agarose)	Agarose %; buffer system
2	Assemble rig (TAE)	Rig; running buffer
3	Prepare samples	Sample identity; loading dye
4	Load ladder + samples	Lane layout; ladder identity
5	Run (100 V, 50 min)	Voltage; run time
6	Stop & retrieve gel	Run complete
7	UV image (gel doc)	Gel image (imager auto-logged)
8	Size bands vs ladder	Observed band sizes
9	Score per lane	Expected vs observed; pass/fail
10	Clone call	Downstream call; e-signature

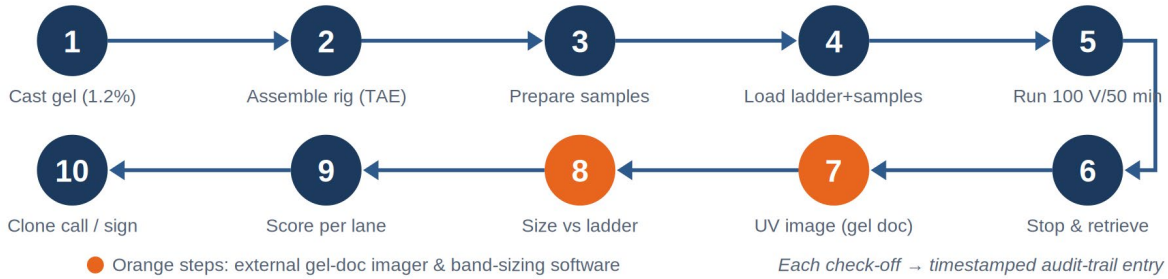

Figure 2. The 10-step agarose gel workflow — orange steps use the external gel-doc imager and band-sizing software; every check-off writes a timestamped entry to the ELabELN audit trail.

© 2026 LabLynx, Inc. — Public information

4. The Use Case: When and How the Template Is Used

The template is intended as a tested baseline for academic and life-sciences research labs running gel electrophoresis. It is configured around a recognizable reference — an EcoRI + BamHI clone screen of pUC19-GFP — but the scenarios it serves span the gel bench and the projects it feeds.

4.1 Restriction-Digest and Clone Screening (Primary Use Case)

The core scenario: running a diagnostic digest across several clones and reading which ones carry the expected fragments. The template captures the lane layout, the expected sizes, the gel image, and the per-lane score in one structured record, so a screen is fully reconstructable — which clone in which lane, which expected bands, which passed — rather than an image filed away from the layout that explains it.

4.2 PCR-Product Verification

A gel confirms that a PCR produced a single product of the right size. Because the expected size and observed band are typed and linked to the run, verification is captured against a documented expectation rather than judged by eye — and, for amplification-centric work, the ELabELN library also offers a PCR template in which the gel is the final verification step.

4.3 Ligation and Cloning Checks

Ligations and assemblies are checked on a gel before committing to a transformation or a prep. Because each check's expected and observed bands are structured and queryable, the decision to proceed is made from the record rather than a remembered impression of the gel.

4.4 RNA Quality Assessment

A gel reveals whether RNA is intact or degraded from the pattern of its ribosomal bands. Because the run parameters and observed pattern are captured as typed fields, an RNA-integrity check becomes a documented, comparable QC record rather than a one-off image.

4.5 Fragment Isolation (Preparative Gels)

A preparative gel separates a fragment for excision and purification. Because the template structures the gel and its bands, moving from an analytical screen to a preparative run is a matter of editing the sample and gel sections, with the excised fragment and its size recorded for the downstream cleanup.

4.6 Teaching, Onboarding, and Multi-PI Collaboration

Gel electrophoresis is a technique every new lab member learns. Because every ELabELN edition includes unlimited users, students, rotating researchers, and collaborators can all work from the same tested baseline, and granular permissions let the PI control who can edit gel runs versus only review them — useful for onboarding and multi-PI collaboration alike.

 © 2026 LabLynx, Inc. — Public information

5. Dependencies and Connected Workflows

The Gel Electrophoresis template does not exist in isolation. It sits inside a network of connected workflows — upstream, parallel, external, and downstream — that give the gel record its full scientific and decision-making value.

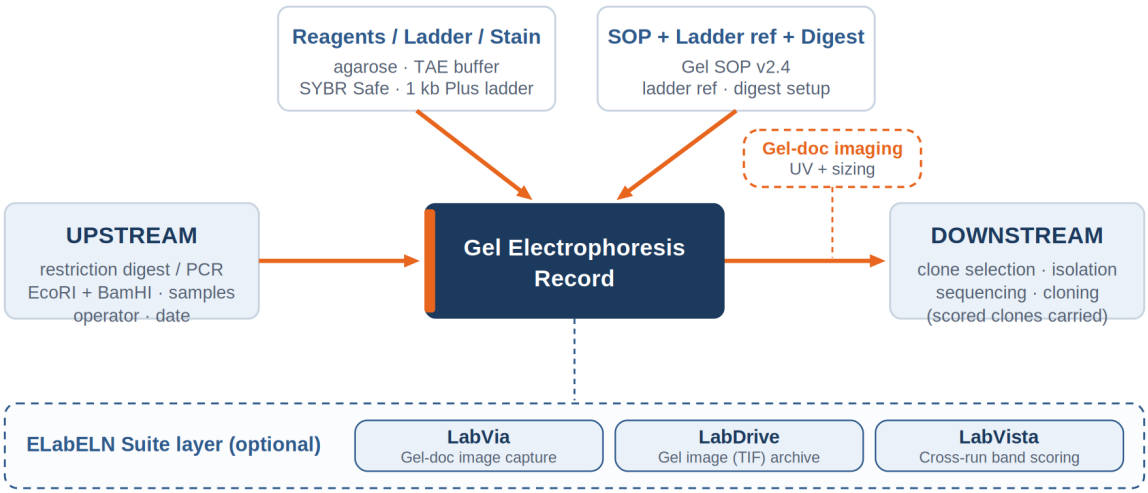

Figure 3. The template connects an upstream digest or PCR workflow, parallel reagent/ladder/stain inventory and SOP/ladder-reference/digest-setup systems, the external gel-doc imager, and downstream clone-selection, isolation, sequencing, and cloning workflows.

5.1 Upstream: The Digest, PCR, or Sample Prep

Every gel begins with something to run — a restriction digest, a PCR product, a ligation, or an RNA sample. The template links to the upstream digest or prep, so opening a gel surfaces the samples, the expected sizes, the operator, and the date — connecting each lane to the reaction that produced it.

5.2 Parallel: Reagent, Ladder, and Stain Inventory

The Compounds section links the agarose, TAE buffer, SYBR Safe stain, DNA ladder, and loading dye directly to the ELabELN inventory. Because ladder identity and stain condition bear on sizing and detection, this traceability is decisive. Through the link the record automatically carries:

- **Lot numbers and recipe references** for the agarose, buffer, ladder, and stain, so a sizing or detection change can be traced to a reagent batch.
- **Stain expiry, shown inline**, so a gel is not imaged with a degraded stain that weakens faint bands.
- **Stock levels**, with low-stock warnings before a run, so a gel is not started against a depleted reagent.

Barcode and QR scanning can populate these links from the physical bottle, keeping lot capture effortless across runs.

© 2026 LabLynx, Inc. — Public information

5.3 Parallel: SOPs, the Ladder Reference, and the Digest Setup

The Links section connects the gel to the lab's Gel Electrophoresis SOP, the ladder reference image, and the digest setup for the run. The digest-setup link is what makes scoring coherent: each gel opens alongside the reaction whose expected fragments it is read against, so the expectation is built in rather than recalled.

5.4 External: The Gel-Doc Imager

Imaging and sizing run on the gel-documentation system outside ELabELN: the stained gel is captured under UV or blue light, and band sizes are read against the ladder, often with the imager's software. ELabELN is the system of record around that instrument: per-lane scoring (clone ID, expected versus observed bands, score, and downstream call) lives in the spreadsheet editor, the gel image (TIF) attaches in the Storage & Files section, and the observed bands and calls are written back into the typed Scoring fields. The image and the interpretation stay together in one record.

5.5 Downstream: Clone Selection, Isolation, Sequencing, and Cloning

The scored gel feeds clone selection, fragment isolation, sequencing, and further cloning — and, because bands and calls are typed and queryable, cross-run and cross-clone comparison. A decision to sequence, isolate, or advance a clone rests on a documented, comparable gel rather than an image recalled from a drive.

5.6 The ELabELN Suite: Instruments and Dashboards

Everything above is available in ELabELN Standard. Labs on the ELabELN Suite gain three layers that connect to the same record: LabVia captures images directly from the gel-doc system and other instruments; LabDrive adds metadata search, versioning, and role-based access across gel-image archives; and LabVista builds cross-run band-pattern and scoring dashboards on the structured gel metadata. The same audit trail and data-integrity controls span all layers.

 © 2026 LabLynx, Inc. — Public information

6. How the Scientist Benefits

The template is built to deliver value at the bench and to the project alike. The benefits compound with every gel a lab runs.

6.1 Start From a Tested Baseline, Not a Rewritten Setup

The scaffold — protocol, typed fields, reagent and ladder links, SOP, and digest-setup link — is already built. The scientist edits a working configuration to match the samples and fragment range they actually resolve, rather than rewriting the gel setup from scratch every run. The first gel is faster, and every subsequent one is consistent and comparable.

6.2 Document by Doing

Checking off each of the 10 steps as it happens produces a contemporaneous, timestamped record as a by-product of the run, and the imager run is auto-logged. There is no separate write-up to reconstruct later.

6.3 Never Lose the Lane Layout or Expected Sizes Again

Because the lane layout and expected sizes are structured fields and the reagents are linked to inventory, the layout, run parameters, ladder, and expectations are captured permanently on every gel. When a band is questioned, the image and its context are on the record — closing the most common gel troubleshooting gap by default.

6.4 Cross-Run Band Patterns You Can Query

Typed run parameters, expected sizes, and per-lane calls make the project queryable. A team can pull every gel of a construct or screen, compare band patterns and scores run by run, and gate on the ladder-referenced sizing at a glance — turning a folder of gel images into a live screening view.

6.5 Per-Lane Scoring and Defensible Result Provenance

Capturing expected versus observed bands and a pass/fail call for every lane makes clone selection a visible, queryable gate rather than a remembered impression — the difference between “the digest looks OK” and a record showing which four of six clones passed and which two had partial digestion. The tamper-evident audit trail, reagent-lot traceability, and electronic signatures give each result a defensible provenance, structured to support cloning-provenance documentation and FDA 21 CFR Part 11 review when the lab’s quality system requires it.

> **At the bench, in practice**
>
> A scientist opens the template, sets the samples and expected sizes to match today’s digest, scans the agarose and stain bottles to capture their lots, and checks off each step as the gel is cast, loaded, run, and imaged. The gel-doc image is attached, the bands are sized against the ladder, and each lane is scored expected-versus-observed with a pass or fail — the per-lane calls confirming which clones carry the right insert. Weeks later, comparing screens, the project lead queries every gel of that construct and sees which clones passed, run by scored run.

© 2026 LabLynx, Inc. — Public information

7. The Gel and Per-Lane Scoring

7.1 The Default Reference: A pUC19-GFP Clone Screen

The template ships configured for a diagnostic EcoRI + BamHI double digest of a pUC19-GFP cloning intermediate, run on a 1.2% agarose gel in TAE with SYBR Safe stain at 100 V for 50 minutes, imaged under UV and sized against a 1 kb Plus DNA Ladder. The 8-lane layout places the ladder, six candidate clones, and an undigested control side by side; a correct clone releases the expected two fragments — the backbone and the insert — while a partial or failed digest shows an extra uncut band. This concrete reference demonstrates the full cast-to-scoring workflow out of the box; swap the samples, agarose percentage, buffer, stain, or ladder to match what your lab runs.

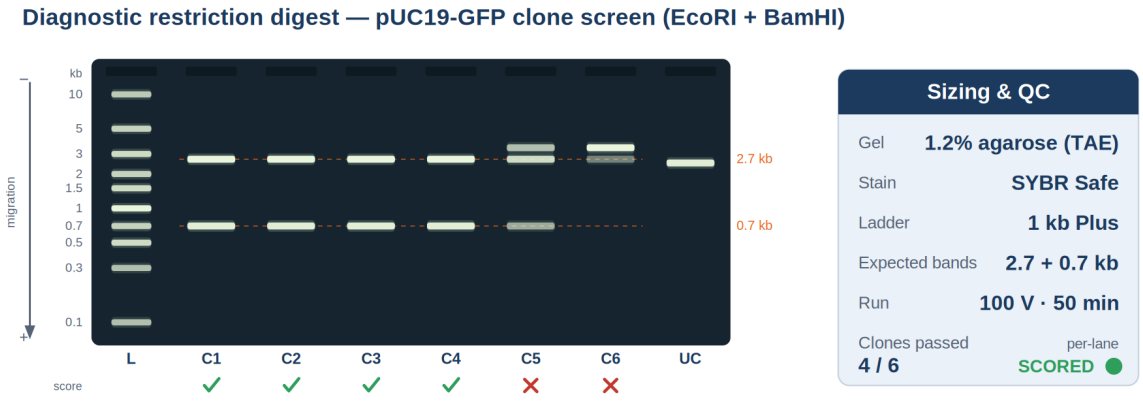

Figure 4. An annotated agarose clone-screening gel — a 1 kb Plus ladder, six EcoRI + BamHI clone digests against the expected 2.7 and 0.7 kb bands, and an undigested control — with the per-lane pass/fail scores and the sizing-and-QC panel. Illustrative; the gel image is captured in ELabELN, and expected versus observed bands and per-lane calls are recorded as typed fields.

 © 2026 LabLynx, Inc. — Public information

7.2 How the Template Handles the Gel, Sizing, and Scoring

The per-lane scoring — clone ID, expected versus observed bands, score, and downstream call — lives in the spreadsheet editor, structured for direct cross-clone selection. The gel image (TIF) attaches in the Storage & Files section, and the observed bands and pass/fail calls are written back into the typed Scoring fields. Because these values are structured and queryable, cross-run comparison is straightforward: query every gel of a construct, gate on the ladder-referenced sizing, and review which clones passed side by side — the screening view a cloning project runs on. Per-lane structured scoring is the difference between “the digest looks OK” and a queryable record showing which clones passed and which had partial digestion.

Element	Default configuration	Editable to
Agarose %	1.2% (routine)	0.7% (>5 kb), 2–3% (<500 bp)
Buffer system	TAE	TBE (higher resolution)
Stain	SYBR Safe	GelRed, EtBr
Run	100 V, 50 min	Any voltage / run time
Ladder	1 kb Plus DNA Ladder	Any DNA / RNA ladder
Application	Analytical clone screen	Preparative; PCR / ligation / RNA QC

Figure 5. The template’s default configuration — fully editable for any agarose percentage, buffer, stain, ladder, or application.

© 2026 LabLynx, Inc. — Public information

8. Configuring the Template for Your Gel

Adapting the template to a different fragment range or application is a matter of editing the relevant sections — not rebuilding the structure.

8.1 Set the Agarose Percentage and Buffer

Choose the agarose percentage for your fragment range — 0.7% for fragments above 5 kb, around 1% for routine work, and 2–3% for fragments below 500 bp — and select the buffer system by editing the Gel section: TAE for routine runs, TBE for higher resolution of small or close fragments.

8.2 Change the Stain and Run Conditions

Switch the stain — SYBR Safe, GelRed, or EtBr — and adjust the voltage and run time by editing the Run section. The lane-layout and scoring fields apply regardless of the stain or run conditions.

8.3 Set the Lane Layout, Ladder, and Expected Sizes

Define the lane layout, choose the DNA or RNA ladder, and record the expected fragment sizes for each sample by editing the Scoring section, so every observed band is scored against a documented expectation rather than judged by eye.

8.4 Move From Analytical to Preparative, and Save a Version

Move from an analytical screen to a preparative fragment isolation by editing the sample and gel sections, then save your edits as a private template scoped to your lab, or publish back to the ELabELN template library. Every subsequent gel opens from your configured baseline, and separate versions can be maintained for different applications or fragment ranges.

© 2026 LabLynx, Inc. — Public information

9. Getting Started

ELabELN Standard is a cloud-hosted SaaS deployment typically live within one to two business days of account creation. There is no hardware to buy, no IT project, and no per-seat licensing.

1. **Start an ELabELN account.** Visit elabeln.com/get-started. ELabELN Standard runs on LabLynx's managed cloud infrastructure.
2. **Provision your instance.** LabLynx provisions the instance within one to two business days; you receive a URL, admin credentials, and onboarding documentation.
3. **Load the Gel Electrophoresis template.** From the template library, select the Gel Electrophoresis (Agarose) template and click "Use This Template." An editable instance opens immediately.
4. **Add your reagents, ladder, and stain to the inventory.** Enter your agarose, buffers, DNA ladder, stain, and loading dye with lot numbers and storage locations. This is a one-time setup step.
5. **Link your SOP, ladder reference, and digest setup.** Upload your Gel Electrophoresis SOP and ladder reference image, and connect the digest or PCR setup to the template's Links section.
6. **Customize the template for your gel.** Edit the agarose percentage, buffer, stain, voltage, ladder, lane layout, and expected sizes, then save as a private team version.
7. **Run your first gel.** Open your saved template, complete the 10-step checklist, attach the gel image, score each lane expected-versus-observed, and sign off.

Typical timeline

Day 1: Account requested at elabeln.com/get-started. Day 2–3: Instance provisioned; reagent, ladder, and stain inventory populated; SOP, ladder reference, and digest setup linked. Day 3–4: Template configured and saved as a private version for your samples and fragment range. Day 4–5: First gel documented in ELabELN with the image attached and every lane scored, captured with an e-signature.

9.1 ELabELN Standard vs. ELabELN Suite

Capability	Standard	Suite
Gel Electrophoresis template access	Included	Included
Unlimited users	Included	Included
Audit trail and e-signatures	Included	Included
Reagent / ladder / stain inventory with lot tracking	Included	Included
SOP, ladder-reference & digest-setup linking	Included	Included
Per-lane scoring table + gel image (TIF) attachments	Included	Included
LabVia direct gel-doc image capture	—	Suite layer
LabDrive gel-image archive search & versioning	—	Suite layer
LabVista cross-run band-pattern & scoring dashboards	—	Suite layer

© 2026 LabLynx, Inc. — Public information

10. Frequently Asked Questions

Can I configure the template for my samples and fragment range?

Yes. Every section is editable. Adjust the agarose percentage (0.7% for fragments above 5 kb, around 1% for routine work, 2–3% for fragments below 500 bp), swap the buffer system (TAE for routine, TBE for higher resolution), change the stain (SYBR Safe, GelRed, EtBr), adjust voltage and run time, or move from analytical to preparative by editing the gel and sample sections. Save your edits as a private template scoped to your lab, or publish back to the ELabELN template library.

How does the template differ from the PCR template's gel verification?

The PCR template includes gel verification as the final step of an amplification workflow. This template treats gel electrophoresis as the primary readout for a broader range of applications — restriction-digest screening, ligation checks, RNA quality assessment, and fragment isolation. Use the PCR template when the gel is downstream of an amplification; use this template for standalone gel analysis.

Does the template handle per-lane scoring across clones?

Yes. The spreadsheet editor captures clone ID, expected versus observed bands per lane, and a pass/fail score, supporting clone-selection workflows. Per-lane structured scoring is the difference between "the digest looks OK" and a queryable record showing which four of six clones passed and which two had partial digestion.

Is it included in every ELabELN plan?

Yes. Every published template in the ELabELN library is available to all subscribers in both the Standard and Suite editions, with unlimited users in either edition and no per-template fee.

Can my whole lab use this template?

Yes. Every ELabELN edition includes unlimited users, so the PI, postdocs, grad students, rotating researchers, and visiting collaborators can all use the template without per-seat charges. Granular permissions let the PI control who can edit gel runs versus only review them.

How quickly can my lab start running gels in ELabELN?

ELabELN Standard cloud deployment is typically live in one to two business days. Once the instance is provisioned, the template loads from the library in a single click, with the agarose percentage, buffer, voltage, and ladder editable to match the gels your lab actually runs.

© 2026 LabLynx, Inc. — Public information

11. Conclusion

The ELabELN Gel Electrophoresis (Agarose) Experiment Template is not simply a digital copy of a gel-doc image. It is a structured, audit-ready, searchable record that connects to your reagent, ladder, and stain inventory, your SOPs and ladder references, the digest or PCR the gel followed, your gel-doc imager, and the downstream selection, sequencing, and cloning workflows. It turns a routine gel into a record whose lanes, expected sizes, bands, and calls are captured, comparable, and defensible.

For a laboratory that runs gels constantly, the benefit compounds with every project. Lane layouts, band patterns, and per-clone calls accumulate into one queryable screening history, and every reagent lot is traceable. Every lane is scored against a documented expectation — so a band judged “about right” by eye can no longer quietly drive a decision, and the comparison that drives clone selection is built in rather than reconstructed from images.

The template is available to all ELabELN subscribers today — a new instance is live in one to two business days, with the first structured gel typically documented within a week of account creation.

> **Get started**
> Visit elabeln.com/get-started or contact LabLynx at 866.522.5969 / info@lablynx.com. Load the Gel Electrophoresis template, configure it for your samples, and run your next gel from a tested, audit-ready baseline.

 © 2026 LabLynx, Inc. — Public information